

Inducible chitinolytic system of *Aspergillus fumigatus*

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Incubation of *Aspergillus fumigatus* NCPF 2140 in growth medium containing 1% chitin as sole carbon source led to induction of specific extracellular chitinolytic activity of 1.5 $\mu\text{mol GlcNAc released min}^{-1}$ (mg protein) $^{-1}$. The effect was repressed by the inclusion of GlcNAc in the medium, indicating regulation by a negative feedback mechanism. Extracellular chitinase activity was inhibited by allosamidin (IC_{50} 0.12 μM). Multiple chitinolytic enzymes were detected on zymograms of extracellular preparations; levels of individual enzymes induced were dependent upon whether cells were incubated with purified colloidal chitin or a crude preparation of crystalline chitin. A major, inducible, 45 kDa chitinase was purified using ammonium sulphate precipitation, chitin affinity chromatography and a novel procedure involving the electroelution of the enzyme from a substrate gel containing glycol chitin. The enzyme is a glycoprotein with endochitinase activity.

Keywords: *Aspergillus fumigatus*, extracellular chitinase, enzyme induction, glycoprotein

INTRODUCTION

Chitin, the linear homopolymer of *N*-acetylglucosamine (GlcNAc), is an important structural component of the crustacean integument, insect exoskeleton and fungal cell wall. The abundance of this polysaccharide in nature is matched by the wide range of organisms that produce chitinolytic enzymes. Plants produce chitinases and other lytic enzymes with roles in defence against fungal pathogens, while bacteria and other microorganisms secrete chitinases that appear to have roles in pathogenesis and/or nutrition (Gooday, 1990; Flach *et al.*, 1992; Sahai & Manocha, 1993; Graham & Sticklen, 1994; Gooday, 1996). Chitinases with nutritional roles release more readily accessible oligomers of GlcNAc from insoluble, crystalline chitin in the environment. Organisms that contain structural chitin produce chitinases with roles during morphogenesis. For example, insect chitinases appear to have crucial roles during ecdysis (Flach *et al.*, 1992) while fungal chitinases have roles during cell separation in yeasts, and are thought to contribute to a number of morphogenetic processes in filamentous fungi including spore germi-

nation, side branch formation and autolysis (Gooday *et al.*, 1992).

Aspergillus fumigatus is a ubiquitous saprophyte and an important opportunistic fungal pathogen of humans (Cohen, 1991; Barnes & Denning, 1993; Campbell, 1994; Lewis *et al.*, 1994). *A. fumigatus* is an interesting example of an organism that appears to produce chitinases with both morphogenetic and nutritional roles. A range of intracellular chitinolytic enzymes and, in particular, chitinases closely associated with cell wall fractions, are thought to have roles during morphogenesis (Hearn *et al.*, 1996, 1997) while *A. fumigatus* represents one of the most abundant known sources of extracellular chitinase (Monreal & Reese, 1969). This would imply that the fungus secretes large quantities of chitinase through a cell wall containing chitin. Currently, the mechanism that enables secretion of chitinase without digestion of the fungal wall is not understood. Similarly, although chitin has been shown to induce the release of chitinases by *A. fumigatus* and a number of other eukaryotes and prokaryotes (Monreal & Reese, 1969; Flach *et al.*, 1992; Sahai & Manocha, 1993; Graham & Sticklen, 1994; Adams & Escott, 1997) there is a paucity of information concerning the mechanism that enables fungi, bacteria and higher plants to detect chitin polymers in the environment. *A. fumigatus* is an ideal model for the investigation of these phenomena and the aim of the current study was to characterize

Abbreviations: GlcNAc, *N*-acetylglucosamine; 4-MU-GlcNAc, 4-methylumbelliferyl-*N*-acetyl- β -D-glucosaminide; 4-MU-(GlcNAc)₂, 4-methylumbelliferyl- β -D-*N,N'*-diacetylchitobioside; 4-MU-(GlcNAc)₃, 4-methylumbelliferyl- β -D-*N,N',N''*-triacetylchitotrioside.

more fully the inducible chitinolytic system of this organism.

METHODS

Chemicals. Unless stated otherwise, all chemicals and reagents were purchased from either Merck (Analar grade) or Sigma-Aldrich. Radiochemicals were supplied by Amersham International. Allosamidin was kindly provided by Lilly Research Laboratories.

Organisms and culture conditions. Spore suspensions of *A. fumigatus* NCPF 2140 in 0.1% Tween 80 obtained from 2–3 d cultures on Sabouraud's agar plates were used to inoculate flasks containing a volume of Vogel's N medium corresponding to one-fifth of the total capacity of the flask (Vogel, 1964) to a final concentration of 1×10^6 spores ml⁻¹. Cultures were supplemented with either sucrose or 1% chitin in the presence or absence of 0.5% GlcNAc. Two chitin preparations were used: commercially available crystalline chitin isolated from crab shells (Sigma) or colloidal chitin prepared as described below. Cultures were incubated in an orbital incubator at 37 °C and 200 r.p.m.

Synthesis of colloidal chitin. Colloidal chitin was synthesized by the acetylation of chitosan according to the method of Molano *et al.* (1977). Following homogenization, the chitin was filtered, washed and ball-milled as described by Dickinson *et al.* (1989). Chitin was resuspended in an appropriate volume and buffer according to its subsequent use.

Protein estimation. Protein concentration was determined using the method of Bradford (1976).

Chitinase assay. Chitinase was assayed using the method of Dickinson *et al.* (1989) with the following modifications. Reaction mixtures contained tritiated chitin suspension [60 µl; 2.6 mg ml⁻¹, 70 kBq (mg chitin)⁻¹], 50 mM Bis Tris/HCl, pH 6.5 (assay buffer), and enzyme preparation (total volume 180 µl) and were incubated at 37 °C in a shaking water bath (160 r.p.m.) for 1 h. The reaction was stopped by the addition of 180 µl 10% trichloroacetic acid. All assays were performed at least in triplicate. Variation between replicates was never greater than 15%.

Glycol chitin zymograms. Glycol chitin was synthesized as described by Trudel & Asselin (1989) and incorporated into polyacrylamide gels using the method of St Leger *et al.* (1993) with the following modifications. Polyacrylamide separating gels (12%) were used with stacking gels of 4.5% polyacrylamide; both contained 0.5% SDS. Samples were boiled for 3 min in a 1:1 dilution with boiling mix [10% stacking gel buffer (0.5% SDS, 0.5 M Tris/HCl, pH 6.8), 2% SDS, 10%, v/v, glycerol, 0.005% bromophenol blue]. Low-molecular-mass protein markers (Pharmacia Biotech) were used to calibrate the gel. Electrophoresis was carried out at 30 mA for approximately 3.5 h using Hoeffer SE 600 electrophoresis apparatus. After SDS-PAGE, molecular mass markers were trimmed off and placed in fixative (methanol/water/acetic acid, 5:5:1, by vol.) and protein bands were visualized by silver staining as described by Hitchcock *et al.* (1989). Chitinolytic activity in the remainder of the gel was detected as described by Escott & Adams (1995).

Purification of 45 kDa chitinase. Proteins were precipitated from culture filtrate (7 d incubation of *A. fumigatus* NCPF 2140 in Vogel's N medium with 1% crystalline chitin as the sole carbon source) using (NH₄)₂SO₄ (60% saturation). Insoluble material was pelleted by centrifugation, resuspended

in 50 mM Bis Tris/HCl, pH 6.5 and dialysed against the same buffer overnight at 4 °C to give a final volume of 10–20 ml (8–15 mg protein). The dialysate was subjected to chitin affinity chromatography essentially as described by Mellor *et al.* (1994) but with the following modifications: the chitin suspension comprised 17 mg (dry wt) colloidal chitin ml⁻¹; the column was eluted against gravity at a flow rate of not more than 1 ml min⁻¹ and bound protein was eluted using 80 mM acetic acid (25 ml; 2 ml fractions).

Electrophoresis and electroelution. Proteins eluted from the chitin affinity matrix were electrophoresed on polyacrylamide gels containing glycol chitin (15 tracks, approx. 120 µl per track) and chitinolytic activity detected essentially as described by Escott & Adams (1995), except that the gel was incubated in 100 mM sodium acetate, pH 4.8, at 30 °C for only 2 h. When the gel was illuminated with UV light, a zone of hydrolysis corresponding to the 45 kDa chitinase was identified readily and excised from the gel using a scalpel. Protein was eluted from the gel by the method of Findlay *et al.* (1989) with the following modifications. The strip of gel was placed in a dialysis tubing bag containing electroelution buffer (50 mM Tris/acetate, pH 7.8, 0.1% SDS), excess buffer was squeezed from the bag and the bag was sealed and placed on the platform of an electrophoresis tank (GNA-200; Pharmacia Biotech) filled with electroelution buffer. Electrophoresis was performed overnight at room temperature and 50 mA. The gel slice was removed from the bag, which was resealed and the contents dialysed overnight at 4 °C against three changes of 50 mM Bis Tris/HCl, pH 6.5, prior to concentration using Centricon 10 devices according to the manufacturer's instructions (Amicon). The presence in the dialysate of a protein of 45 kDa was confirmed following SDS-PAGE using the PhastSystem (Pharmacia Biotech) and silver staining of gels, although low-molecular-mass breakdown products were frequently detected following electroelution.

Protein sequencing. The electroeluted 45 kDa chitinase was electrophoresed using SDS-PAGE under non-reducing conditions as described above for glycol chitin zymograms except that glycol chitin was omitted from gels (1.5 µg protein per lane). The gel was blotted using semi-dry blotting apparatus (Novoblot; Pharmacia Biotech) with CAPS transfer buffer (1 mM CAPS, 20% methanol, pH 11). The transferred protein was visualized by staining with sulphorhodamine B (0.005%, in 30% methanol, 0.2% acetic acid) for at least 2 min before rinsing briefly in distilled water and air drying. The sections of blot containing the 45 kDa chitinase were excised and stored at -20 °C prior to sequencing. The N-terminus of the 45 kDa chitinase was sequenced using automated solid-phase microsequencing (kindly performed by Dr A. Moir, University of Sheffield, UK).

Overlay gels incorporating 4-methylumbelliferyl-chitooligomer substrates. SDS-PAGE was performed as described above for glycol chitin zymograms except that glycol chitin was omitted from the separating gel and Mini-Protein II gel electrophoresis apparatus (Bio-Rad) was used. Samples were prepared in non-reducing boiling mix and electrophoresed at 200 V for approximately 45 min. Molecular mass markers were trimmed from the gel and placed in fixative. The remainder of the gel was incubated in casein buffer (Escott & Adams, 1995). The gel was incubated for 30 min in 100 mM sodium acetate, pH 4.8, with one change of buffer and placed onto a glass plate for support. The overlay gel was as described by Pan *et al.* (1991) except that instead of glycol chitin, 4-methylumbelliferyl-N-acetyl-β-D-glucosaminide (4-MU-GlcNAc), 4-methylumbelliferyl-β-D-N,N'

diacetylchitobioside [4-MU-(GlcNAc)₂] or 4-methylumbelliferyl- β -D-N,N',N''-triacetylchitotrioside [4-MU-(GlcNAc)₃] (0.03%) was used as substrate (Tronsmo & Harman, 1993). Overlay gels were allowed to polymerize at room temperature for at least 1.5 h and the gels were incubated together for 15 min at 37 °C in a moist atmosphere. Illumination with UV identified fluorescent bands where enzyme activity had released 4-methylumbelliferone from the chitooligomer.

Detection of glycoproteins. The periodic acid-Schiff method was used to visualize glycoproteins following SDS-PAGE (Dewald *et al.*, 1974). Glycoproteins stained pink on a clear background.

RESULTS

Induction of extracellular chitinase

Chitinase was detected in the culture filtrate 72 h after *A. fumigatus* was inoculated into Vogel's N medium containing 1% colloidal chitin as the sole carbon source (Fig. 1). Similar results were obtained when 1% crystalline chitin was used as the carbon source (data not shown). The presence of chitin in the growth medium prevented accurate determination of growth phase based on estimation of biomass. However, it was evident from microscopic examination of cultures that the initial secretion of chitinase coincided with the onset of stationary phase, while the highest levels of extracellular chitinase were detected with the onset of autolysis. A very high level of specific chitinase activity [$1.5 \mu\text{mol GlcNAc released min}^{-1} (\text{mg protein})^{-1}$] was detected at this time. Under the conditions described, chitinase activity was not detected when sucrose was included in the culture medium; identical results were obtained in the presence or absence of chitin. However, chitinase activity was detected in a freeze-dried concentrate of

culture medium from *A. fumigatus* NCPF 2140 grown for 22 h in Vogel's N medium with sucrose as the sole carbon source (results not shown). This suggests that the constitutive secretion of chitinase occurs at such a low level that it is normally beyond the sensitivity of the assay method employed during the present study. When 0.5% GlcNAc was included in culture medium containing chitin as the sole carbon source, the level of extracellular chitinase produced was reduced markedly but the level of extracellular protein was unaffected (Table 1).

Crystalline and colloidal chitin appeared to induce a different range of chitinases (Fig. 2) although most of the chitinases induced by colloidal chitin were also induced by commercial chitin. There were also apparent differences in the relative activities of the enzymes induced by each chitin preparation. This was particularly noticeable for the triplet enzymes with molecular masses of approximately 32, 28 and 24 kDa. The apparent approximate molecular masses of the major extracellular chitinases induced by colloidal chitin were 110, 69, 45, 32, 28 and 24 kDa while those induced by crystalline chitin were of approximate molecular masses 110, 94, 69, 62, 45, 39, 32, 28, 24 and 20 kDa. These experiments were repeated several times with the same results. The instability of proteinase inhibitors such as PMSF made it difficult to determine whether any of the low-molecular-mass chitinases detected on zymograms were attributable to proteolysis of larger enzymes during lengthy incubations of up to 7 d. However, when chitinases were isolated from *A. fumigatus* mycelia a range of enzymes of differing molecular mass, similar to that detected during the present study, was identified in the presence or absence of aspartate-, cysteine-, metallo- or serine-proteinase inhibitors (Hearn *et al.*, 1997), suggesting that proteolysis does not contribute to the wide range of enzymes detected in cellular and extracellular extracts. GlcNAc (0.5%) did not seem to affect the range of chitinases induced by either crystalline or colloidal chitin; instead, the level of each chitinase induced by the chitin preparations was reduced in the presence of the monomer. When a commercial preparation of *N*-acetylglucosaminidase (3 $\mu\text{g protein}$) was electrophoresed in a glycol chitin gel, no chitinolytic activity was detected.

The chitinolytic activity detected in culture filtrates was clearly attributable to extracellular chitinases, as the specific chitinase inhibitor allosamidin was a potent inhibitor of chitinolysis. Inhibition curves for extracellular chitinases induced by colloidal or commercial chitin indicated IC₅₀ values of approximately 0.1 μM (Fig. 3).

Purification and characterization of an extracellular chitinase

An extracellular 45 kDa chitinase was purified by ammonium sulphate precipitation, chitin affinity chromatography and electroelution from a polyacrylamide gel containing glycol chitin. Analysis of the N-

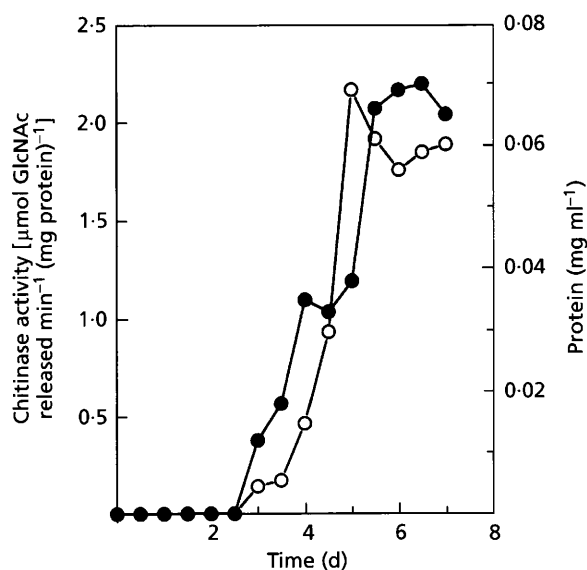


Fig. 1. Induction of extracellular chitinase of *A. fumigatus* cultured in a growth medium containing 1% colloidal chitin. ○, Chitinase activity; ●, total protein. The experiment was repeated with the same results.

Table 1. Effect of GlcNAc on the induction of extracellular chitinase following inclusion of chitin in the growth medium and incubation for 4 or 7 d

The experiment was repeated with essentially the same result.

Addition(s) to growth medium*	4 d		7 d	
	Total protein ($\mu\text{g ml}^{-1}$)	Specific chitinase activity†	Total protein ($\mu\text{g ml}^{-1}$)	Specific chitinase activity†
Colloidal chitin	39	0.200	99	1.251
Colloidal chitin, GlcNAc	59	0.103	107	0.452
Commercial chitin	40	0.908	54	1.296
Commercial chitin, GlcNAc	42	0.458	46	0.509

* Vogel's N medium (Vogel, 1964) was used throughout. Colloidal or commercial chitin was used at 1%. GlcNAc was used at 0.5%.

† $\mu\text{mol GlcNAc released min}^{-1} (\text{mg protein})^{-1}$.

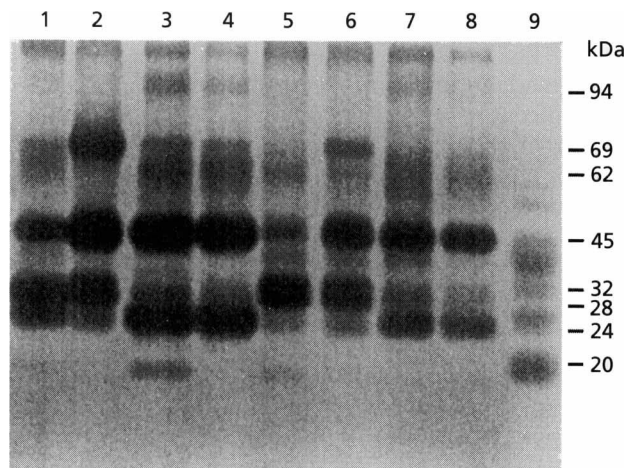


Fig. 2. Zymogram demonstrating induction of extracellular chitinases of *A. fumigatus* and the effect of GlcNAc on induction. The culture medium was Vogel's N medium supplemented with different chitin preparations (1%). The polyacrylamide gel (12% acrylamide) incorporated glycol chitin as a substrate for chitinase; 1.25 μg protein was used in lanes 1–8 and 3 μg protein in lane 9. Lanes: 1 and 2, colloidal chitin, harvested at 4 d and 7 d, respectively; 3 and 4, commercial chitin, harvested at 4 d and 7 d, respectively; 5 and 6, colloidal chitin and 0.5% GlcNAc harvested at 4 d and 7 d, respectively; 7 and 8, commercial chitin and 0.5% GlcNAc harvested at 4 d and 7 d, respectively. Lane 9, *Streptomyces griseus* chitinase (Sigma).

terminus of this protein revealed homology with chitinases from other fungi and most notably with the enzymes from *Coccidioides immitis*, *Trichoderma harzianum* and *Aphanocladium album* (Fig. 4). PCR primers based on the sequence of the N-terminus of the *Aspergillus fumigatus* chitinase and conserved residues

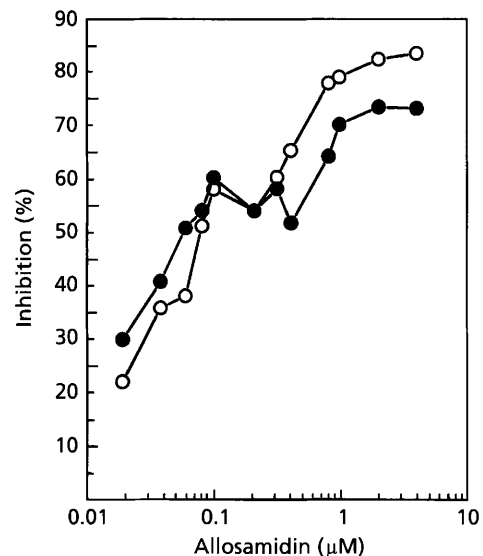


Fig. 3. Effect of allosamidin on extracellular chitinase induced by growth in medium containing colloidal (○) or crystalline (●) chitin as sole carbon source. The experiment was repeated with essentially the same result.

in fungal chitinases were used to amplify two fragments of approximately 450 bp from *A. fumigatus*. The fragments have >60% homology with the *C. immitis* *cts1* gene (A. K. Jaques, G. M. Escott, R. Barton, C. A. Hitchcock & D. J. Adams, unpublished).

When the purified 45 kDa chitinase was electrophoresed in an SDS-polyacrylamide gel and the gel incubated with an overlay gel containing 4-MU-GlcNAc, 4-MU-(GlcNAc)₂ or 4-MU-(GlcNAc)₃, the chitinase did not cleave 4-MU-GlcNAc (data not shown) but released 4-

	1	15 16	30 31	45 46	60
<i>A. fumigatus</i>				SSGYRSVVYF	VNWAIYGRN
<i>C. immitis</i>	-MRFLIGALLTLQTL	VQASSMSSMPNYPV	PEAPAEGGFRSVVYF	VNWAIYGRGHNPQDL	
<i>T. harzianum</i>	MLGFLGKSVALLAAL	QATLISASPVTANDV	SVEKRASGYANAVYF	TNWGIYGRNFQPQNL	
<i>A. album</i>	MLSFVKKSIALVAAL	QAVTALATPISSE-A	GVEKRGSGFANAVYF	TNWGIYGRNFQPADL	

Fig. 4. Homology between the N-terminus of the *A. fumigatus* 45 kDa chitinase and the N-termini of chitinases from other fungi. Homology was identified by submitting the *A. fumigatus* sequence and all fungal chitinase protein sequences (24 sequences) currently lodged in GenBank to the Multiple Sequence Alignment facility of the BCM Search Launcher (Human Genome Center, Houston, TX, USA). The proteins showing the greatest homology with the *A. fumigatus* N-terminus are shown here. GenBank accession numbers: *Coccidioides immitis* complement fixation antigen-chitinase, U60807; *Trichoderma harzianum* ech42, X79381; *Aphanocladium album* chi1, P32470.

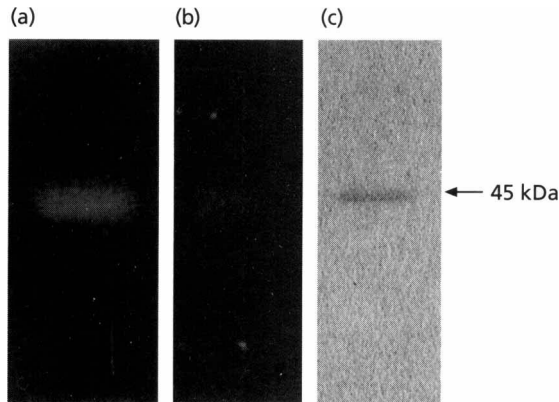


Fig. 5. Characterization of the 45 kDa chitinase in polyacrylamide gels. Detection of chitinolytic activity in a polyacrylamide gel (12% acrylamide, containing 1.25 µg protein per track) used an overlay gel of polyacrylamide containing (a) 4MU-(GlcNAc)₃, or (b) 4MU-(GlcNAc)₂ as substrate. The experiment was repeated twice with the same results. (c) Periodic acid-Schiff-stained polyacrylamide gel (12% acrylamide, containing 2 µg protein).

methylumbelliferone from 4-MU-(GlcNAc)₂ and 4-MU-(GlcNAc)₃ (Fig. 5a, b). These results indicate that the 45 kDa enzyme is an endochitinase; an exochitinase would cleave 4-MU-(GlcNAc)₃ at the central β,1-4 bond, yielding 4-MU-GlcNAc and chitobiose and, therefore, no fluorescent product would be produced. The enzyme also cleaved the longer chain chitooligomer substrate more readily (Fig. 5a, b), a phenomenon observed previously for other endochitinases (Cabib, 1987; Tronsmo *et al.*, 1996). A single protein band corresponding to the band with endochitinase activity was detected when gels were silver stained (results not shown).

When ConA-Sepharose was used for lectin affinity chromatography, most of the extracellular chitinases adhered to the column matrix (results not shown), indicating that these enzymes are glycoproteins. Confirmation that the purified 45 kDa chitinase is a glycoprotein was obtained by staining the chitinase in an SDS-polyacrylamide gel using the periodic acid-Schiff method (Fig. 5c).

DISCUSSION

Our results indicate clearly that under certain growth conditions, *A. fumigatus* secretes abundant quantities of chitinase and the array of extracellular chitinolytic enzymes detected appears to reflect the wide range of chitinases identified in intracellular, cell wall and cell surface preparations from this organism (Hearn *et al.*, 1996, 1997). The application of a procedure we described previously for the purification of chitinase from *Candida albicans* (Mellor *et al.*, 1994), combined with the electroelution of protein from a substrate gel containing glycol chitin, enabled the rapid purification of an extracellular chitinase from *A. fumigatus*. The enzyme is a glycoprotein with endochitinase activity. Determination of the N-terminal sequence of this protein, coupled with the identification of regions of conserved sequence in chitinases from a range of organisms, has enabled us to design PCR primers which we have used for the identification of at least three chitinase genes in *A. fumigatus* (A. K. Jaques, G. M. Escott, R. Barton, C. A. Hitchcock & D. J. Adams, unpublished). The results of the present study suggest that *A. fumigatus* contains several genes encoding chitinases. In the longer term, gene disruption experiments will allow us to determine precise roles for these enzymes. Fungal chitinases with roles during differentiation have been proposed as potential targets for antifungal agents. Comparison of the roles of *A. fumigatus* chitinases with those of chitinolytic enzymes produced by human cells (Overdijk & Van Steijn, 1994; Escott & Adams, 1995; Renkema *et al.*, 1995) should enable a more thorough evaluation of the potential of fungal chitinases as targets for antifungal agents. If human chitinases are shown to have roles in host defence, it may be necessary to reconsider the suitability of fungal chitinases as drug targets. Instead, the identification of mechanisms that regulate the activity of microbial chitinases and other autolysins at the level of transcription or post-translational modification may unveil alternative, specific targets for novel antifungal drugs.

It is equally important that mechanisms regulating the induction and secretion of chitinases from *A. fumigatus* should be characterized more fully. A wide range of organisms including higher plants, bacteria and a number of fungi produce abundant quantities of chi-

tinase in response to chitinous materials in the environment. However, it is not yet clear how these organisms detect the presence of the insoluble chitin polymer outside the cell. The induction of differing profiles of *A. fumigatus* chitinolytic enzymes by semi-synthetic (colloidal) and crude (crab shell) chitin preparations is of particular interest. Thus, for example, a chitinase of apparent molecular mass 94 kDa was induced when *A. fumigatus* was cultured in a medium containing colloidal chitin as the sole carbon source (Fig. 2). However, this enzyme was not detected when *A. fumigatus* was grown in a medium containing crystalline chitin. In addition, comparison of tracks 1 and 2 with tracks 3 and 4 of Fig. 2 indicates clearly that the levels of individual chitinases induced by one chitin preparation differed markedly from the levels of enzymes induced by the other. A similar effect was demonstrated for *Aphanocladium album*, which produced nine chitinases when grown with colloidal chitin as the sole carbon source, but only three chitinases when crystalline chitin was used as the carbon source (Studer *et al.*, 1992). This work and the results obtained during the present study imply that *Aphanocladium album* and *Aspergillus fumigatus* can detect differing configurations of chitin and, perhaps, cross-linking of the polysaccharide to other polymers in the growth medium. Partial degradation of chitinous material in the environment, by low levels of chitinase secreted constitutively by *Aspergillus fumigatus*, will yield a range of oligomers which may stimulate transcription of chitinase genes by interacting with receptors in the fungal cell (Adams & Escott, 1997). This induction mechanism would enable *Aspergillus fumigatus* to utilize a range of complex chitinous substrates while repression of chitinase secretion by GlcNAc prevents wasteful synthesis of the enzyme in the presence of abundant quantities of the ultimate product of chitin hydrolysis. Similar induction-repression mechanisms were identified in *Trichoderma harzianum*, where both glucose and GlcNAc repressed the synthesis of induced chitinase (Ulhoa & Peberdy, 1991), and in *Metarhizium anisopliae*, where chitinase synthesis was repressed by the addition of alanine, a readily utilized nutrient source (St Leger *et al.*, 1993). The extensive chitinolytic system of *A. fumigatus* provides an interesting model for further studies of chitinase induction and repression, phenomena that are important in plant defence, entomo- and myco-parasitism and the production of bacterial and fungal chitinases that enable biodegradation of chitin in the environment.

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