

Nucleotide sequence of shallot virus X RNA reveals a 5'-proximal cistron closely related to those of potexviruses and a unique arrangement of the 3'-proximal cistrons

K. V. Kanyuka, V. K. Vishnichenko, K. E. Levay, D. Yu. Kondrikov, E. V. Ryabov and S. K. Zavriev*

Institute of Agricultural Biotechnology, 12 Pskovskaya Street, Moscow 127253, Russia

The 8890 nucleotide RNA sequence of shallot virus X (ShVX), a new virus isolated from shallot, has been determined. The sequence contains six open reading frames (ORFs) which encode putative proteins (in the 5' to 3' direction) of M_r 194528 (ORF1), 26333 (ORF2), 11245 (ORF3), 42209 (ORF4), 28486 (ORF5) and 14741 (ORF6). The ORF1 protein was found to be highly homologous to the putative potexvirus RNA replicases; ORF2, -3, -5 and -6 proteins also have analogues among the potex- and/or

carlavirus-encoded proteins. ORF3 is followed by an AUG-lacking frame coding for an amino acid sequence homologous to that of the 7K to 8K proteins of the triple gene block of the above-mentioned viruses. The putative ORF4 protein has no reliable homology with proteins in the database. The results obtained testify that, except for the unique 42K protein gene, the ShVX genome combines a number of elements typical of both carla- and potexviruses.

Introduction

We have recently found a new kind of flexuous filamentous potyvirus-like particle in shallot plants (Vishnichenko *et al.*, 1992) which is, however, serologically unrelated to either of two identified *Allium* potyviruses, onion yellow dwarf virus and leek yellow stripe virus, and forms no cytoplasmic inclusions typical of potyvirus infection (Hollings & Brunt, 1981). This previously unknown viral pathogen of shallot has been provisionally named 'shallot virus X' (ShVX) (Vishnichenko *et al.*, 1992). The RNA isolated from ShVX preparations is an ssRNA about 9000 nucleotides long.

In this report we present the nucleotide sequence of ShVX RNA. Analysis of open reading frames (ORFs) and the amino acid sequences of the putative protein products reveals a combination of elements typical of both carla- and potexviruses. An exception to this is a putative ORF4 protein, which bears no analogy to any protein known to date.

Methods

Virus purification and preparation of viral RNA. The virus was isolated from shallot plants (selection sample 83 from the Institute of Vegetable and Seed Production, Moscow Region, Russia) grown at 28 °C as described previously (Vishnichenko *et al.*, 1992). The virus suspension was mixed with an equal volume of extraction buffer containing 0.2 M-ammonium carbonate pH 9.0, 2 mM-EDTA, 2% SDS and proteinase K

(15 µg/mg of virus). The mixture was incubated for 15 min at room temperature, after which the RNA was extracted twice with phenol-chloroform, once with chloroform and precipitated with 2.5 volumes of cold ethanol in the presence of 0.1 M-sodium acetate. The RNA precipitate was washed with 70% ethanol, dissolved in sterile water and stored frozen at -70 °C.

cDNA synthesis, cloning and sequencing. The double-stranded cDNA for the ShVX RNA was synthesized essentially as described previously (Levay & Zavriev, 1991). The cDNA was ligated to a *Sma*I-digested pGEM-7Zf(+) (Promega) or *Hinc*II-digested pGEM-3Z plasmid and transformed into competent *Escherichia coli* XL-1B cells (Stratagene Gene Cloning System). To detect clones containing the 3'-terminal virus-specific sequences, ampicillin-resistant transformants were screened by filter colony hybridization with short fragments of the ³²P-labelled first strand of ShVX cDNA produced with oligo(dT)₁₅-primed reverse transcriptase. The arrangement and the insert size of the selected recombinant cDNA clones were determined by cross-hybridization and restriction analysis of plasmid DNA.

A nested series of exonuclease III deletions were generated from the original cDNA clones by use of the Erase-a-Base system (Promega). Dideoxynucleotide sequencing of ss- and dsDNA templates was carried out using [α -³²P]dATP (Institute of Applied Chemistry, St Petersburg, Russia) and modified T7 DNA polymerase (Sequenase, U.S. Biochemicals). The extreme 5'-terminal nucleotides of ShVX RNA were sequenced by dideoxynucleotide sequencing using reverse transcriptase and the ³²P-end-labelled synthetic primer 5' dCAGCTTCGTGTTCCGGG 3' complementary to nucleotides 137 to 153 of the final sequence. The cDNA was sequenced in both directions and about 80% of the sequence was determined by examining two or three clones covering each particular region.

In vitro translation. For *in vitro* translation of the ShVX ORF2, -4 and -5 proteins, the RNAs were synthesized with phage T7 or SP6 RNA polymerase from linearized plasmids pX55 (cDNA insert 5229 to 6135, T7), pX5 (5500 to 7745, T7) and pX34 (7430 to 8890, SP6), respectively.

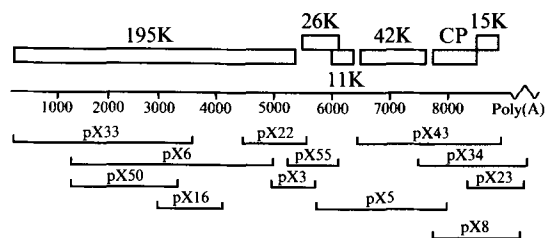


Fig. 1. Schematic representation of six ORFs in ShVX RNA and long overlapping cDNA clones used for sequencing. All are shown relative to their location in the ShVX genome.

Each RNA (1 µg) was translated in a mixture of 10 µCi of [³⁵S]methionine, 125 µM of each amino acid except methionine and 10 µl of rabbit reticulocyte lysate (25 µl final volume). The translation products were analysed according to Laemmli (1970) in 15% polyacrylamide-SDS gels.

Isolation and Northern blot hybridization of the RNA from infected plants. Total RNA was isolated from green shallot tissue as described by Palmiter (1974). Denaturing gel electrophoresis of total RNA was carried out in a formaldehyde-containing 1% agarose gel, after which the nucleic acid was transferred onto a 0.45 µm charge-modified nylon membrane filter (Sigma) in 20 × SSC using an LKB apparatus for vacuum blotting of nucleic acids, as described by Kroczek & Siebert (1990). Virus-specific RNAs were revealed with a heavily ³²P-labelled RNA transcript complementary to the 3'-proximal region of ShVX RNA which was synthesized using phage SP6 RNA polymerase on the pX23 plasmid (cDNA insert 8336 to 8890; Fig. 1).

Virus sequences and calculation of protein similarities. The amino acid sequences of the proteins of potato virus S (PVS; MacKenzie *et al.*, 1989), helenium virus S (HelVS; Foster *et al.*, 1990), lily symptomless virus (LSV; Memelink *et al.*, 1990), carnation latent virus (CLV; Haylor *et al.*, 1990), potato virus M (PVM; Zavriev *et al.*, 1991), chrysanthemum virus B (CVB; Levay & Zavriev, 1991), potato aucuba mosaic virus (PAMV; Bundin *et al.*, 1986), potato virus X (PVX; Huisman *et al.*, 1988), white clover mosaic virus (WCIMV; Forster *et al.*, 1988), narcissus mosaic virus (NMV; Zuidema *et al.*, 1989), papaya mosaic virus (PMV; Sit *et al.*, 1989), lily virus X (LVX; Memelink *et al.*, 1990), clover yellow mosaic virus (CYMV; Sit *et al.*, 1990), strawberry mild yellow edge-associated potyvirus (SMYEV; Jelkmann *et al.*, 1990), turnip yellow mosaic virus (TYMV; Morch *et al.*, 1988) and apple chlorotic leaf spot virus (ACLSV; German *et al.*, 1990) were predicted from the nucleotide sequences of the viral RNAs. The initial sequence alignments of the virus proteins were made with the MULTALIN program (Corpet, 1988). The detailed alignments were made upon visual inspection of the homologous regions revealed using the program.

Results and Discussion

Isolation and sequencing of viral cDNA clones

To elucidate the nucleotide sequence of ShVX RNA, four cDNA clones were initially selected: pX23, pX8, pX34 and pX43 (Fig. 1). The cDNA inserts of these clones except pX43 contained a poly(A) tail 15 to 43 nucleotides long, with identical sequences adjacent to it. Clone pX43 overlapped with clones pX23, pX8 and

pX34, and ended 71 nucleotides away from the poly(A) tail.

Clones containing the 5'-terminal regions of ShVX RNA were selected stepwise by colony hybridization with 3'-end-³²P-labelled cDNA inserts containing sequences most remote from the RNA 3' end. In this way we picked out another eight plasmids that contained overlapping cDNA inserts (Fig. 1).

The cDNA nucleotide sequence corresponding to the ShVX RNA [8890 nucleotides excluding the 3'-terminal poly(A)] and the deduced polypeptide sequences are shown in Fig. 2. Since the first nucleotide could not be unequivocally identified by dideoxynucleotide sequencing using reverse transcriptase (see Methods), it is denoted N in Fig. 2. The sequence of the ShVX 5'-terminal 98 nucleotide non-translated region is very similar to that of PVX RNA determined by Morozov *et al.* (1983) and Huisman *et al.* (1988). The 3'-terminal non-translated region is 112 nucleotides long and ends in a poly(A) tail. The ShVX RNA also contains internal non-coding regions between ORF1 and ORF2 (62 nucleotides), between ORF3 and ORF4 (113 nucleotides) and between ORF4 and ORF5 (26 nucleotides). The overall base composition of ShVX RNA is 28.6% A, 22.2% U, 20.6% G and 28.6% C.

The ShVX RNA sequence determined with independent cDNA clones displays a fairly high incidence of base changes (about 4%) which mostly, however, do not result in amino acid changes (Fig. 2).

Coding regions and genome organization

Analysis of the ShVX RNA sequence shows the presence of six ORFs (Fig. 1). ORF1 (positions 99 to 5252) encodes a 1718 residue polypeptide of M_r 194 528 (195K), ORF2 (positions 5315 to 6037) encodes a 241 residue polypeptide of M_r 26 333 (26K), ORF3 (positions 6018 to 6326) encodes a 103 residue polypeptide of M_r 11 245 (11K), ORF4 (positions 6440 to 7579) encodes a 380 residue polypeptide of M_r 42 209 (42K), ORF5 (positions 7606 to 8777) encodes a 262 residue polypeptide of M_r 28 486 (28K) and ORF6 (positions 8394 to 8777) encodes a 128 residue polypeptide of M_r 14 741 (15K). Analysis of the negative strand sequence of the ShVX RNA reveals a single ORF for a putative 12K protein in the region complementary to ORF1 (positions 1702 to 1342 on the positive strand).

Protein sequence analysis and comparison with other plant virus-encoded proteins

(i) ORF1

The ShVX ORF1 protein sequence displays a high extent of similarity in three extensive regions with virus-

NACTCAACAAACACACACACACAAACGACAAACTTGACTGAGTGTGATAA 60
HTAVQKL
GGAAATCAGGAAGCCGTGTACACCCATCAAGCAAGTATGACTGCTGTGAAAACCTT 120
FDQISDPNNTKAGVSNACFEA
TTGACCAATATGCGACCEGAACGAAGTGTGCTTCCAAATGCTGTGTTGAAGCGG 180
AQRPRPKAMAIAPFVSVTPE
CGCACCGGCTCAAGAAGCTATGGCTATAGCCCTTTTCGCTCACCCAGAGG 240
ALTLERFGITTSPTTSTHT
CTCTCAGCTTGGAGCTTGGCATCACCCTCCCTTTGCGCCACCACTCACACAC 300
HAADKJIENDCLTIIGHVLP
ATGCTGCCACAAATAATGGAATGACTGCTCAAAATATTTGGCCACTATGTACCA 360
KREAYTLIQLKRL
AACGABAGGAGTCACTTATCAACTCAAGCGAGCAAAATCTCTTGGGAGG 420
QPSQDNHFQNYCHEPQDVLRY
ACCAGTCAAGCAATTTCCAAATATTTGCGCAGGCAAGGACCACTTAGGTAGC 480
GITHPHNSCPVVAATEAVLAD
GGATCCACCCCAACTTTCGCGAGTGTAAACAGAAATACGCTTTGTGGCGGATA 540
TLHFMSRPLLYLHLSRNPKL
CCTTACCTTTATGCTCCAGACGCTGACCACTTATGACGAGCAATCCAAAGCTAG 600
ERL FATLVLPIEAORHLPSSL
ABGCGCTTTCGCGAGTACTCACTCAATTAAGGCTCAGCAGGCTACCGAGCTTAT 660
FPDVRLEYYKDFHFAVHPGG
TTCCTGTGTAGACAGTGAATATTAAGACGATTTGCTTATATGCGGAGGCT 720
HGGGAYVHVSYGLTKWLDWTA
ACGGTGGCGTCTTACGACTTCTTACGACTCTCAAAATGGCTAGACACGCTCAAG 780
VGPVDTKSSITNHPCTD
TGSGCCSTAGATTAACATACTCAACCACTTACGCTTACGCTTACGCTTACG 840
LSIEKIETKAAHHIMFIQRT
TTAGCATAGAAAGATGAACCAAGCAGGCGCACTATAAATGTTACACAGCGACAC 900
RAQVDWPLPPIWVHASEYV
GGGCTCAGGTGATGGCCACTGCCGCCATATGGCTCAGCAGGCTCAGATAGCTCA 960
KLPFLFIYPPEANVQKAYSPHT
AATACCCCTCATTTCTACCCGAGAGGCAATACAGAGGACTACCCGACAGCT 1020
LIKRNQLYCTFSVKAVSLRDI
TCATCAAGAGATGCAACTGTATGCTTCAAGCAAGGCTATCACTAAGAGACACT 1080
FAKLRQVIEITAGELVRYSMAD
TTGCTAGCTTGGCAGCTTATGAGACACAAAGACTTGTGCTACTTACTGGCAGAC 1140
LIRLANVFLFITGMNQVSDY
TCATGAGCTAGCAGCTACTTCTGCTTCACTACGGGTATGATCAAGTACGAGCT 1200
ESPLLEHLFQKMHKASIRHRL
AATCTCACTGCTGAGAAATTTATTCGGGAAATGCTGTCTCAATCGGCTAGACTCA 1260
RTFFQNLGKTSYAAALLTVT
GAATCTTCCAAAACCTGCTGGCAAAATATGCTGCGCTTCTCAGGCTCAGG 1320
DVIPVHTTTPKREAVGEL
ATGCTATCCGGCCACACTACCCAGCCCAAGCAGAGAGGCTCTGGGCGAGTAT 1380
WFQEPKWSVSTHTQPRKEHH
GGTTTCAGAGCCCAATGAGCGTGAAGCAGTGAACCAACCCGCAAGAAACATCACC 1440
RLQMTVTLWLLAFHQLSESSG
GTCTCAGATGACTGAGCTTGTCTAGCGGTTTACCACATGTAGAGTTCGGGCTCA 1500
MSEPCNHSESTPQRTATSOQ
TGTGAGAAAGCTTAATATTCGGATCCAGCCAGCAGCAGCAGCAGCAGCAGCAG 1560
KAAKLTTSQKHNRRTDQTH
AAGCGCCAGTAAACGACTCAGCAAGACACAGCAGCAGCAGCAGCAGCAGCAG 1620
NPQYPLMLTIAPHMRRHSL
ACCACAACTGCTCCGCTGATGTTGAGATGTGCCGATGATGCTCGACATTCGTA 1680
MKKTIATP CRTLEEISLDL
TGAAGAGACGATTTGCACTCATGCCGGGCTGCAAGAGATTTGAGCTTTGACTTAG 1740
DDFDLDPNEASNEPPSANEQ
ATGATTTGATGACTGCTCAAGCAAGCTTCAAGCAAGCAGCAGCAGCAGCAGCAG 1800
SPDNHAETTRGVFPCECGT
CCCCGCAATCATGCTGAAACACCCAGSAGSAGTGTTCCTGGAGTGGGCGACCG 1860
EITVMSFGR AIEVAGVNLTD
AAATCAGACTTACTCTTTGCTGCGATAGAAAGTTCAGGCGTAATCTGACGACC 1920
HNKGRLLAFYSRDRGQGYSYT
ACATGAAGGAGGCTGCTGCTTCTACTCAGSAGTGGCAGGCTACTTACTACTG 1980
GYSHKKSQGWLEGLDKLIEAC
GCTACTCCCAAAATCGAAGGCTGATGAGGCTTGTGACAAGCTGATGCAAGATGCG 2040
GKAPTTVNLQCLVYDQGSR
GTGAAAACCTACCTTACACAGCTGCTGTTACAGAAATACGAGCAGGCTCAAGAA 2100
IGFHSDEQA IYPKGNKILTV
TAGGTTTCCATAGGACGCAAGCTATATACCCAAAAGGCAATAAATCTCAGCTGTA 2160
NAGSGTGF I KCAKGETG
AGCAGCGGCTCCGCAATTTGGCTAATGATGTGCAAAAGGAGAAACCACTGATC 2220
LEDGDYFQMPSSGFEHTKHHN
TGGAGATGAGTATTTCCAAATGCGAAGGGTTTCCAGBAAATCTCAGGACATAAGC 2280
VYAVTPRLSFTFRSTVYNSQ
TCGTGAGTACACTGCTTACCTTCACTTCCAGTCTGACGCTGATGCAAGTACCAA 2340
KKPAPEPEKLNQHNACPKPSO
AGAAACCCGCAAGCTGAGAGCTGATCAAAACATGCTGCTTCCAAAACCTCAGAT 2400
PSNAGSKQHKKTHPAKGNK
CATCAAGGCTAGCGGCAAGCAGCAAGAAACCCCTGCAAGGCAAGCAAGAT 2460

SSSPNLEPLDAPTYEILKHL
CTTCTTCAACAACTCGAACCCCTGGAGCCTACTGCTGAGATCTCAAGCTCCAGC 2520
GFTALTPHQHOGTQJIRPVYF
GCTTCACTGCCCTCACTCCCAACTGAGCGCAGCTGCAAACTGCGCCTGTTATTTCA 2580
NKDIHLRKRKAVKTDHSPPAR
ACAAGAGATCTTACGAGGCAAGAGCAGTGAAGCTGACATGCTCCACCACAGCGC 2640
PFFDLATSLHRGIYTHKIDM
CATCTTTGACTAGTACTACTCTCCACCGAGCATTATACGCAAAATGCAACACC 2700
RRATAYSDVKNHLLTGLVLP
GTAGACTACGGCTACATGCTGACTCAGAAATACCTCACTGCTTAGTCTCCCTA 2760
KLDROD LSSMVALAETTTRE
AATGGACGCTGACTTGTGAGTCTTGGTAGCGCTGCTGAGACTCACACAGCGAAG 2820
YAVLAIHGAGGAGGAGKRSALQE
YTGCGCTTACGCTTACTGAGCAGGAGGAGCTGGTAAAGCTGAGCTACAGGAAC 2880
LLRSSPELAD SINIVVPTIN
TGTGATGCTTCCCGACAGCTGCGACAGCACTAATCTGGTCAACCTATAAACC 2940
LANDWKAKLPDAPRRVMTF
TCGCTAACGACTGAAAGCCAAATACCAAAAGGAGCTCGTAGAGTCACTGCTCC 3000
AAAAGCTTGTGAGGAGAGTGCACACTGAGCAGTATTTGACGATTCAGGCAACTTC 3060
PAGFYDAYLAIKVNVLEAIL
CTGCAGATTTGTGGACGCTACTGCTATCAGGTGAAGCTGAGCTGGCGATACGA 3120
TGDQDRQSTHHERESQISSL
CTGGATCAAGCCCACTCAACCTATCAAGATACAGGCTGAATCTCAAAATGCTACTTC 3180
QSNIAOFSKYADYVLNATHR
AAGTAACTGCCCACTTCAAGTACGAGATTTACTCTCAATGCCACTATAGGC 3240
QPRRLANIKVHAEROLGGA
AGCCAGGCGACTGCTAACCTTATCAAGATACAGGCTGAGCGCAATGGGGGGGCG 3300
VLKANI V PDLAMVLPVAFRS
TGCTCAAAAGCAATGTGCGAGATCTGGCAGTGTGCTGCGCTGATTCGCGACCC 3360
QSLTTDLGRHAMTYAGCQGL
AGTCACTTTACGAGCTTACGCGACAGCAGCTGACTACCGCGCTGTCAGGGCTCA 3420
TLNHLLTILDKDTP LSCDEV
CACTTAATCAGTACGATGCTATGAGCAAGGACCCCTTTATGCTGAGGAGGTC 3480
LYTAFSRASESITFVNTHSD
TCTACAGGCAATTTTCCGCTGCTGAGTCEAATGCTTTGTGAACCCACTTGATA 3540
HPAFLAKD TATPYLKTLSH
ACCGTCTTCTGGCAAACTGATGCAACCCCACTCAAGACTGATGATGGG 3600
VTRGAGGATGAGGAGCTGAGGAGCTGCGGCTTCCGCTTACGAGCTTGTAAAGATG 3660
VPTKTHIPVANDKVLQLEGI
TGCCCACTAAGCACATATACCGTACGCAATGCAAGGCTGCAACTGAGGGGAAGTGC 3720
EAMEDKDTRELWSGEEKTNL
AGCCCATGAGCAAGGACAGCCGATGCTGAGGAGAGAAAGAAACCACTGA 3780
MGTQCTGAGGCTTGTGAGCTTCTCCACCAACAGCAAGCAGGAGGAGCT 3840
LFKITIGERIRHATPEQNAK
GTTCAAAATACAAATGAGGAGCATTGATGCAAGCCTGAGCAAAACGTAAGC 3900
QLRHTLHAGD LFEAYAFH
AGCTGAGCACACACTCAATCCGCGGATCTACTTTGAGGCTGACGACAGTTCATGA 3960
KYVKE TQFPDKRLWTHCRQL
AAGTCCCAAGAAAGCAGCCCTTTGACAACGTTTATGCAACTTGGCTCAACTAG 4020
ALR TYL SKPTSSNLQGGARD
CTCTGCGACTTACTCTGCAAGCTACACTTACCTTCAAGAGGGGCGAGCAGCAGC 4080
PDPFDNAIALFNKSNQVVKLL
CTGATTTCCAGCAATGCGCTAGCCTAATTAACAACTGCAATGGTCAAGAACTTG 4140
EKVGARFKAGQTAISAFKQEV
AGAAAGTGGGCTGCTTCAAGCTGGCAAGCATATGAGCTTTAAAGCAAGAGTGG 4200
VLLTTNHALRKRKREHQHP
TACTTCAACAGGACTTGGCACTTACTTCAAGAAAGAAAGAGGACCAAGCCGCG 4260
DNVFINCERTEPEQNAFVMT
ACAAGCTTTTATATGTTGAAAGACCCCTGCAAACTCAAGCCTTTGATGACTA 4320
KWD RPNHYTS DYTQYDQSQ
AGTGGACTTTCAGCGCAACTACACTTGGCAGTACACAAATGCAACTCTCAG 4380
DAAFLNFEIRKARHLGVPE D
ACGCGCTTCTTACTTCAATGAAAGAGGCGACTTATGAGTGTGCCGGAAGATG 4440
IMRLSAEGPTFDANTECNIA
TATGCTTCAAGCGGCAAGGCGCAACTTGAAGCTTCAAGCAACAGAGTCAATTCAG 4500
YDALRFR L G D V R A S Y A G D
ATGACCCCTCAGATTCAGCTTGGAGAGGCTTGGGCTGCTAGCGCGGTGACCGC 4620

LVRDKACEERAGWVYSESLF
TGGTAAGAGCAAGGATGCAAGAGCGCGCGGCTGGGTTTATCTGAGAGTTTATCA 4680
SLKAKPLVTKNPKDFCGWRLT
GCTCAAAAGTAAACCACTGCTCAAAACCACTGACTTCTGGCGTGGCAGCTACCA 4740
RHGIVKSPIDLYQSLQALR
GACATGCTTCAATGCGCCACTCACTTACCAATGCTGCTGACTGCGACTCAGAC 4800
LGKIDEVKRSY AIDYLFAYR
TTGAAAATTTGACGAGGTAAGCGAGTATGCTGACTACTGTTTGGCTACCGCC 4860
LGDKIYDIFDELEKHLV
TGGGCAAAATATAGCACTTCTGAGCAGGAGCACTAGAAAACATCAAGTTGGTA 4920
TRTLIKKGMQPPESGNHLP
CTCGACCTCAAGAAAGGTAAGCTGACTTCACTGCTGAGCGGCAAGCCTTCAAT 4980
FHTTIRLIDPDAVVKQSY
TCCACTTACTGCTGATGATGAGAGTCCAGAGCAGTGAAGTCCAGCTATCG 5040
ECDRILLKQPHIIDDYIPAG
AGTGGCAGGACTTACTGAGGACGACACACTTATGATGATTAATCTGCTGCGCA 5100
TQPRNTEHPSADARDMTRA
CTCAACCCGTAACGAGGACCCAGCTTGCAGATGCTGAGACATGACGAGCGGT 5160
CNLSAEGEKLAFEGGNTNHLFR
GCAATCTAGGCAAAAGTTCCTTGGGGAAGCAACAACTCAACCTTATTTCGAA 5220
P P L S
T S W E G R S P L S H *
CATCTTGGGAGGCTTCCCTTCTGCAACTTGTCAACCACTAAATTAACCAACC 5280
M K T D L L L L Q
CCTTAGCATTAGTATGTTTATAGGTGTTGAAATGAGAGCTGACTTCACTACAA 5340
L S N H N F R T S E P I K E P L I I H
ACTATCAACAATAATTTACAGCAGCTGAGAGCGATCAAGAGCCTCTCATATACA 5400
G V P G S G K S T L Y R A L V T Y R S T
CGGTGCTGCTGGTCCGCAATTCACCTTATGAGGAGCAGTACTGATTCAGCAGTCA 5460
V A C T L G A P Y G S H L A G P P G V T
TTCGCTGACACTTGGAGACTTACGGGCAAGTACGATTCAGGAGGCTGATC 5520
P G L T Q S L T D H E T R I L D E Y Q L
TCTGGGTCACAACTCTGACAGCAGCAGGAGCAGAACTTGCAGGAGTACAGGTT 5580
G T E S D L K P F N V L V G D P F Q G
AGGCAGGATTCGACTGAAACCTTCAAGCTTGTGAGGCGATCACTTCAAGGTA 5640
L H L K A H Y K S F S H R V P R I J C
CTGCACTCAAGGCCCACTAGCTGAGGAGTTCACATCTGATCAACAAAGATCTTG 5700
N F L Q S L G Y E I A G S K P G E L A Q
CACTTCTACAGCTCAGGTCAGAAATGCGGGATCAAAACCGGCGAGCTGCGACA 5760
L P I Y G P N P S G P T G Q V L H L G P
ACTACCAATATACGGCCCAACCTTGTGGCAACCGGCGAAGCTCTACCTAGGAC 5820
L S R R L T Q S H G V C S K L P S E V Q
TCTCCCGAGGCTCACTCAAAAGCGGTTGCTGCTCAAACTCCCTCAAGTTCA 5880
G L E F E V T L V Y H S S E F E R N R
AGGACTGAGTGAAGAGTCACTCTGATGACTCTCTGATTTGAGGCGAAGC 5940
V G F Y I A T A R L G R L N L I D A R D
CGTAGTTTCTACATTCGCGCACTGAGGCTTGGAGCACTAATCTTACTACCGGAC 6000
M S F A P P P D Y S K I Y L
T L E I P H E L C P T S *
AACATGGAGTCCCACTGAGTTTGGCCCACTGACTACTCAAGATTTACTAG 6060
A L G C L G L G L G F V V Y A S R V N H L
CCCTAGGCTTGGGCTGCTGCGGATGTTGCTACGCTTCTGGGTCAGCAGCTAC 6120
P H V G D A C T H N L P H G G Q Y C D G N
CACAGTGGCAGCACTCAACTTCTGCCACAGGAGGCGAGTACTGAGCGCAACA 6180
K R V L Y S G P K S G S S P T N H L W P
AGCCTGCTTACTCAGGCAACCACTTGTGATCAGCAGCAACCAACCTGCGGCT 6240
F I T V I L T L A I L L T S C P R R
TCATTACGTTATGCTCCTCCGCTGATGCTTCACTAGTGGCTCGCCGCGGTG 6300
V C I R C S D H *
TTTGATACGCTGCTTCAACTATTAAGGAGGACTCAAGGTGCTTATTAACCT 6360
ACAGGCTCCACCCAGATTTCAACTGCTGAGGAGCACTGCTGAGTATTAAG 6420
N V I V T F H I A R D
TCATCGATGAGCAAGCTGTAATGTCAGCACTTCCACTTGAAGCAGCAGGAG 6480
R I N C V K D V R H I V T N Q V P A
CCGCTCAATATGTTGAAAGCTGAGGAGTATGCTAATCAAGCTGCTGCTGCT 6540
T R K L G S I E T T L E T T I G
CACTCGGAATGGGTCATAGAGCAGCAGCAAGGAAATTCAGACAGAAACCTG 6600
G T T I S D C V S L L R N L R S E T T
TGGTTCAGAACCACTCCGACTGCTGCTACTCAGGATTTAAGATGCGAGACT 6660
R N F H T L L S R N T A E P T G Q A O T Q
CGCAGCTTCAACACTTACGCGGCTGAGCAACCAAGGCAAGCAGCAACCA 6720
L R Q G F D E P D G H K S E Q R T F S
ACTGCTGAGGCTTGTGAGGAGCAGGCGCAAGGAGGAGCAAGGATTTCTTCT 6780
N I D T A L M A T Q A L L N H V P P A R
AAATTTGATAGCCCTGAAAGTCAACAGAGGCTTCTTAAACAGCTGACCCGCTAG 6840

Y T L P P A P L P V H E S F G Q L H A L
 ATACAGCTACCTGACGACCGTGGCCGTTAATGATCGTTGGACAATTGACGCGCTT 6900
 H L N T L E W L T H I N H N L D S M L N
 GCACCTGAACACATTGGAATGBCGACGACATCAATCAACTTGACCTCCATGCTTAA 6960
 M L N P A N L M S O G T P L S R L K D A
 CATGCTCAACCTCGCAATCAATGTCGCAAGGACTGCACTCAGCGCTTTGAAGAGCC 7020
 V R T L T Q N N H T I Q S D Q Q K I L A
 AGTCCCTACTCTCACAAAACATGAATATCTCAGTGGATCAGCAGAGATTCTAGC 7080
 S T S A T N H S D I L R K L E S L D T G
 GTCCACTTCAGCTACTAACCCTCAGACATCTGCGCAAGTGGAAATCATTAGATACAGG 7140
 L K Q L G I R L D V V V S S L N H M S E
 CCTAAGCACTCGGAATCAGCTAGAGTGTGTGTGCTCAAGCTCAACACATGAGCGA 7200
 R P P T P S H D T A S S S T S T D P N P
 ACGACCGCCCTCCTTGCATGACACCGCAAGCTCATCCACATCAACAGACCCAAATCC 7260
 L P P Y Q A G V H P H T C R T Y G H E L
 GTTACCACCATACCAAGGCTTCCACCGTCTTCTGCGTACTATGCGAATATCTCT 7320
 Y N G I D S R I P H D V T G R P A S T S
 CTACACGGTACTGACTCTCGCATTCGGATGATGATGCGGCGCCGACGACGACTTC 7380
 L K L T I T V E C S E Q N T R V M F T L
 CCTTAAATTAACATTACTGTTGATGATGCGAACAACCAAGGATTAACCTCAGCGT 7440
 L D D E G I L L S D S I E T K H K L Q H
 ACTTGACATGGATACCTGCTGCTGATCTGAGACCAACACAACTTCAACA 7500
 I P S D C L S L I H A R C P K F V Y K F
 CATCCCAAGCGATTGTTTATCTTAATACATGCCAGATGCCCAAAATTTGTTTACAAT 7560
 R G E G L C * M N E E D
 TAGGGGTGAAGTCTGTGTTAAGTTGTCGGGTAGCTCGAAATACAATGAACGAAGGAT 7620
 L N R L N A S G D L N G A N N Q R I P A
 TTGAATGTTGATGATGATGATGATGATGATGATGATGATGATGATGATGATGATGAT 7680
 G P S G V N Q P I P S V Y A G G Q N Q F
 GGACCATCGCGCTCAACCAAGGCTCCGAGGCTGACGACGAGTGGTCAAAATCAAT 7740
 R P S G G L G N Q G S R P T E S S N Q D
 AGACCTCGGAGGACTTGGCAACCAAGCTCAGCCCACTGAACTGAGTAACTCAGGAT 7800
 E L L P T E A E I E A I T S D V E S N S
 GAATATTGCCCTGAAGCTGAGATTGAAGCTCAGTACGCGAGCTGGATCCAATTC 7860
 V A P K A T I R E I L D T L Q A K R Q N
 GTAGCACCACAAAGCACCATTCCGCAATCTGGACCTTCAACCAATGATGATGATGAT 7920
 A T P K D L F S L A W A C Y H N G S S R
 GCCACCCGAAAGTACTTCTTCTTATGCTGAGCTGCTGCTGCTGCTGCTGCTGCTGCT 7980
 F V N L M T D A P C G I T H A D L K T L
 TTCGTGAATCTGAATGATGATGATGATGATGATGATGATGATGATGATGATGATGAT 8040
 W K A S A T L R Q F C S Y A K S C Y V
 TGAAGGCTTTCGCGACGCTCAGACAGTCTGCGACCTACGCTAAATCATGCTATGTT 8100
 S G K Q Q K P P A N S R K G Y P E E
 TCAGGGAACAGCAAGAACCTCCTGCTAAGTGTGCTGAAAGGATATCTGTAAGAG 8160
 A K F A G F D F F N A V L S E S S P A P
 GCCAATTTGCGGCTTGAATTTCTCAACGAGCTGATGATGATGATGATGATGATGATGAT 8220
 P G G M R F K P T Q A E I L G H S M N A
 CCGGCGGAAATCGGTTCAACCCACGCAAGCTGAAATCTGCGGCACTCAATGAAGCT 8280
 K N S I V E S R Q S S H N V S T R A D L
 AAATGTCTAGTGTGATGCTCCGCAATCAGTGCATGTTCACTAGAGCGGACCTC 8340
 L G R Q Q I N E Q P K P P H I T F *
 CTAGGCGCCACAGATTACGCAACCCGACGCGCGATGATAACTTCTGATGATGAT 8400
 P H D L N L L C C L H F S K P S L P N D
 CCCAGCTAATCTTCTTCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 8460
 L K T L L F R A C E T S C K L N R R L L
 TCAAACTCTCTTCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 8520
 D N K P F Q G T S K C A K R R R A K R Y
 ACAATAAGCCTTTTCAAGGACCTTAAGTGTGCTAAAGCGCGAGAGCAAAAGCTTATA 8580
 H R C F O C G A Y L Y D D H V C K R F T
 ATAGATGTTTGAATGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 8640
 S R S N S D C L S V I H Q G P A K L Y A
 GTCGCTAAATCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCTGCT 8700
 E G A Y R A N S D A E Q L I M N D M L L
 AAGGGCTTACCGGCAACTCAGACCGCGCAAGCTTAAATTAAGACGACATGCTATTA 8760
 I K S L K L *
 TTAATCTTAAATTAAGGCTTGGCCCAAGCTCCCACTGGGTTACAGGGCTCTGG 8820
 ACGTGAACCAAGCACTCAGCATGTTGATATAGCTAACTACTGCTGCTACACATTTG 8880
 TCCCCGCGACA_n 8900

Fig. 2. The entire nucleotide sequence corresponding to ShVX RNA. The DNA sequence is shown as the equivalent of the viral positive-strand RNA, and the amino acid sequence of six major ORFs is presented above the nucleotide sequence. The first nucleotide is denoted by 'N' because it has not been identified. Variants in the nucleotide sequence (lower case letters) and the corresponding amino acid changes are shown.

(a)

ShVX	50	TLERFGITTSFPATTSHTHAADKIENDCLT-16-LKRSKI- 8-FQNYCHEPKDVLRYG
PVX	50	D**G***A*N*YSIEL****A**T***K**E-15-***PR*- 9-***VAL**R**A**P
PVM	54	K*SMA**V**S**AVV**S**PVC**TL**Y**Y-15-I*ER*-11-VV*RYVTS*A**R**T
ACLSV	54	WFTKS*VYL***V**V**N**S**PGC**TL**H**F-20-N**M**M- 9-IL*RLV**A**K**A**
TYMV	55	L*N**S**P**G**L**G**T**S**H**P**H**T**T**F**C-15-M**P**F- 8-LK*RLH**N**S**T**P

(b)

	(I)	(II)	(III)	(IV)	(V)	(VI)
ShVX	907	VAVLAIHGAGGAGKSRALQELL-13-VPTINLANDWKAL-19-KSVTFDDYGLK-15-				
PVX	727	**ACV***S***H**I**K**A*-12-L**NE**R**L**S**K**V-19-G**I**V***S**A**-				
PVM	1158	L*SLH**V**T**F**S***L**F**K**N*-11-S**R**R**A**E**F**R**T**V-33-G**Q**V**L**E**M**Q**L**Y-17-				
ACLSV	1051	*K**I**Y**G**F**F**A**S***H**I**N**K**I-13-C**R**R**F**K**S**E**G-19-R**L**F**L**E**T**S**L*-22-				
TYMV	968	TP*VHFA*FA*C**T**Y**I**Q**I*-11-C**T**E**R**T**E**T**A**M-20-S**R**L**I**V**I**E**I**Y**M*-17-				
ShVX			AILLTGDQRQSTH-28-ATHRQPRRL-45-AMTYAGCQGLTLNHLTIIL-10-VLYTAFSRAS			
PVX			I***S***V**Y-28-***N**K**D*-46-TF*****K**P**K**V**V*-10-M**M**L**K**T			
PVM			L**F**V**P**A**D**Y-29-R**S**F**L**N**C**N-59-VL*EGEST**F**M**G**Y**I-10-R**M**I**R**F**R			
ACLSV			IVCT**P**L**A**G**Y-27-V**S**Y**I**N**K**F**I-58-V**F**G**E**S**T**F**C**G**V**V*-10-H**I**M**V**I**T**F**R			
TYMV			V**I**L**P**L**G**E**Y-28-W**S**Y**I**Q**C**I-45-S**C**I**S**S**S**F**C**D**P**A**V**V*-11-N**G**L**V**L**T**S**R			

(c)

		A	B	C
ShVX	1416	YTSYDITQYDQSQ-40-IMRLSAEGPTFDANTEC-15-RASYAGDGLVRD-16-		
PVX	1239	L**A**N***A**F***-40-***T**G***-15-AQV***S**L*-16-		
PVM	1692	T**E***A**F**A**A*-40-***F**G**A**S**L**F**L**A-15-F**I**C**F**M**C**A**S-16-		
ACLSV	1585	V**E***A**F**V**A*-40-***F**T**G**P**S**L**F**L**A-15-P**I**C**F**M**C**A**L-16-		
TYMV	1576	I**A**N***A**F***-40-C***T**G**P**G**Y**D**Y-14-P**I**M**V***S**L**I*-16-		
ShVX		SLKAKPL- 6-FCGWRL		
PVX		L**S**V**V-11-***L**I		
PVM		K***V**Q- 6-***H**		
ACLSV		***V**N- 6-***H**		
TYMV		H**R**F**L**E- 6-***V**V		

Fig. 3. Sequence comparison of the three extensive domains carrying the putative methyltransferase (a), NTP-dependent DNA helicase (b) and RNA polymerase (c) activities of the ShVX ORF1 protein with those of potex-, carla- and tymoviruses, and closterovirus ACLSV. Numbers following abbreviations indicate the sequence position of the first compared amino acid. The lengths separating the motifs are indicated. Stars indicate identical amino acids. Domains and individual motifs therein are shown according to Habili & Symons (1989), Poch *et al.* (1989) and Morozov *et al.* (1990a).

specific RNA replicases of potex-, carla- and tymoviruses, as well as the closterovirus ACLSV (Fig. 3). These regions in the N-terminal, central and C-terminal parts of the ShVX 195K protein contain motifs typical of the putative methyltransferase (Fig. 3a), helicase (Fig. 3b) and polymerase (Fig. 3c) domains, respectively (Morozov *et al.*, 1990a; Habili & Symons, 1989; Poch *et al.*, 1989).

The closest similarity is found between the ShVX ORF1 protein and the putative RNA replicases of potexviruses (Fig. 3). It should be mentioned that general homology is observed virtually throughout the entire length of these proteins barring the region between the putative methyltransferase and helicase domains, which is characteristic of all potexviruses (Skryabin *et al.*, 1988; Rozanov *et al.*, 1990). We conclude that the ShVX ORF1 encodes a virus-specific RNA replicase evolutionarily closely related to those of potexviruses.

Table 1. Percentage similarity between ShVX ORF2 and ORF3 proteins and the corresponding proteins of some potex- and carlaviruses

Virus	ORF2	ORF3
PVX	30	36
WCIMV	26	30
CYMV	27	34
NMV	11	39
LVX	16	41
PMV	30	39
PVM	25	37
PVS	25	40
LSV	28	38
CVB	31	40

(ii) ORF2 and ORF3

All carla- and potexviruses whose genome structure has been established to date, except LVX (Memelink *et al.*, 1990), code for proteins of the triple gene block (Morozov *et al.*, 1989). The ORF2 26K polypeptide contains amino acid motifs which are conserved in NTP-dependent DNA helicases, including the GXGKS/T motif (Gorbalenya *et al.*, 1988). The same conserved motifs are also found in homologous proteins of barley stripe mosaic hordeivirus (58K protein), beet necrotic yellow vein furovirus (42K protein), and *Nicotiana velutina* mosaic virus (29K protein) (Rupasov *et al.*, 1989; Randles & Rohde, 1990). Homologies between the ShVX 26K protein and its counterparts of carla- and potexviruses are shown in Table 1. The *in vitro* translation product of the ORF2-containing RNA transcript is shown in Fig. 4, lane 1.

The ShVX RNA ORF3 encodes an 11K protein analogous to the corresponding triple gene block proteins of all carla- and potexviruses (Table 1). Analysis of the hydrophobicity of the 11K protein using the method of Kyte & Doolittle (1982) shows long hydrophobic stretches. This is also typical of the 12K and 7K to 8K proteins encoded by the triple gene blocks of carla- and potexviruses, as well as of small non-virion proteins of some other viruses (Rupasov *et al.*, 1990). The ShVX 11K protein can probably interact with membranes, as demonstrated *in vitro* for the 12K and 8K proteins of PVX and the 7K proteins of PVM and PVS (Morozov *et al.*, 1990b, 1991).

The ShVX RNA also contains an ORF lacking the initiation codon and coding for an amino acid sequence homologous to those of the 7K to 8K proteins of carla- and potexviruses. This ORF is located around the non-translated region between ORF3 and -4 (Fig. 1). The same situation is found in the LVX genome (Memelink *et al.*, 1990). The 7K to 8K protein analogues of ShVX and LVX might be initiated through the use of an unusual

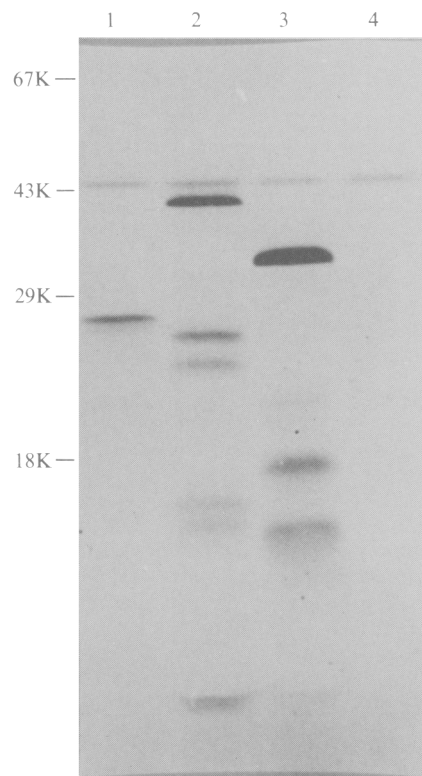


Fig. 4. The ShVX ORF2 (lane 1), ORF4 (lane 2) and ORF5 (lane 3) proteins translated in rabbit reticulocyte lysates. Lane 4, no template (negative control). Positions and size of protein markers (LMW, Pharmacia) are given on the left.

initiation codon, or expressed by some as yet unknown alternative translation mechanism.

(iii) ORF4

The polypeptide encoded by ORF4 attracts most interest, not only because such an ORF is absent from the carla- and potexvirus genomes, but also because no appreciable homology has been found between this protein and those available from the protein sequence database. Another fact worth attention is that this putative polypeptide is extremely rich in serine residues. Analysis of the 42K sequence according to Trifonov's algorithm (Trifonov, 1987) suggests that it can be expressed in eukaryotic systems. Indeed, *in vitro* translation of an ORF4-containing RNA transcript gives rise to a 40K to 42K protein (Fig. 4, lane 2).

(iv) ORF5

The location of ORF5 and the high homology of the encoded polypeptide with the coat proteins of carla- and potexviruses (Fig. 5), testify that this ORF encodes the ShVX coat protein. The ORF5 28K protein migrates in the gel as a 32K to 36K protein (Fig. 4, lane 3), which could be due to the high hydrophilicity evident from its

ShVX	149	ATLRQFCSSYAKSCYVSGKQKPPANWSRKGYPEEAKFAGDFFNAVL
PVX	120	C*****HK**PVVNNMHLTNS***QAQ*FKP*H***A***G*TN
WCTMV	95	C*TI*****F*NTVNIIMLDT*+**K**S*****DG*NH
NMV	95	*IT*****F**VWNLLDSDNV**G*AKQL*DDC*****EG**
SMYEV	129	C***L**MF**PVNKAVDNR**G**NLQFT*P****A***DG*N
CYMV	95	S***R**R**F**VIWNYALRKNQ****ASQN*K*ADR****A***S*
PAMV	133	ISP*****I*VNNMLH-NE*****AKI*FK*DY****A***DA*D*
PMV	96	TS**K**R**F*PIIWWL-RTD*MA****EAS**KPS****A***DG*E*
LVX	90	LP**P**R**F**VWNRLSHDL****ADSQF*A**R**A***DG*TN
PVM	189	E***R**R**L**P*VTWNNHMLTHNA***D**AAM*FQY*DR**A***C*DY*E*
CVB	198	S***K**R**L**AVANNYMHLL*QT**SD**AM*FHPNV**A***C*DY*E*
HeTVS	182	*G**R**R**L**P*VTWNNYMHLLHDS**SD**ASM*FAPNV**A***C*DY*E*
LSV	176	*G**K**R**L**P*VNN*MLVRNQ***D**QAM*FQYNTR**A***T*DY*TN
PVS	178	*G**K**R**L**P*VNNYMLV*NR**SD**QAM*FQWN**A***T*DY*TN
ShVX		ESSPAPPGG-MRFKPTQAEILGHSMNAKMSIV 230
PVX		PAAIM*KE*LI*-P*SE**MNAQT*-AFVK* 200
WCTMV		PAALM*AD*LI*-G*SD****A*QT-***QVAL 175
NMV		PAALD*AD*LI*-P*S*R**QA**T-***YGAL 190
SMYEV		PA*QEV*LRQ---**PQ**YASAT-H*DVAT 207
CYMV		SAALS***LI*-E*SPN*RMANET-***NVHL 175
PAMV		PAALE*SQVVRH---*DK*RAA*GV-V*WASL 211
PMV		PAAMQ**S*LL*-S**E*R*IASAT-***QVHL 175
LVX		SAA*Q**D*LI*-P**EL*LSAAQT-***FAAL 170
PVM		TAAVQ*LE*LI*-R**PR**KVA*NT-H*DIAT 269
CVB		GAAIR*S**IVP*-**R**YVAYNT-Y**LAL 278
HeTVS		PAAVQ*LV*V*IP-R**RD**YVAYNA-Y*LI*LV 262
LSV		QAAIQ*VE*LI*-R**S**V*IA*NA-H*QLAL 256
PVS		GAAIQ*VE*LI*-R**PE**T*IA*NA-H*SMAT 258

Fig. 5. Comparison of the central part of the ShVX coat protein sequence with corresponding regions of the carla- and potexviral coat proteins. Gaps (—) are introduced to increase similarity. Stars indicate identical amino acids. Numbers at the beginning and the end of the sequences indicate the position of the first and last amino acids in the compared regions, respectively.

ShVX	43	GTSKCAKRRRAKRYNRCFDGCGAY-LYDHHVCKRFTSRNSDCLSVIHQGPALKY
PVM	40	*R**Y**R****SIA*CHRC---*R*W*P*P*T*---**T*---CDNKHCY-*GIS*
CVB	39	*R**S**Y**R****ELG*CHRC---*R*V*P*P*L*F---PEIT*---CDNRTCY-*GIS*
PVS	37	*R**T**Y**S**K****R**S**IG*CHRC---*R*V*P*P*---VGN*K---CDNRTCY-*GIS*
LSV	66	*R**R**Y**R****LQIG*CHRC---*R*V*P*P*---VCG*K---CDNKTCY-*GIS*
CLV	40	*E**Y**R****SIA*CHRC---AVSPGF---Y**T*---CDGKTCY-*GLSA*
HeTVS	43	***TY**R****SIL*CHRC---*R*V*P*P*L*---P*SKK---CDNRTCY-*GIS*

Fig. 6. Sequence comparison of the conserved region of the ShVX ORF6 protein with those of the analogous carlaviral proteins. Cys (His) residues that take part in the formation of the putative Zn finger structure are in bold type. Gaps (—) are introduced to increase similarity. Stars indicate identical amino acids.

amino acid sequence. This protein has further been shown to comigrate in the gel with the ShVX virion protein and to react specifically in a Western blot with an antiserum against ShVX (V. K. Vishnichenko and others, unpublished results).

(v) ORF6

The coat protein gene ORF5 is followed by an ORF coding for a cysteine-rich protein. This is a feature typical of all carlaviruses (Haylor *et al.*, 1990; Levay & Zavriev, 1991). The ORF6 protein, like the analogous proteins of other carlaviruses, contains a highly conserved region which comprises a basic arginine-rich domain and a putative Zn finger motif (Klug & Rhodes, 1987). However, one of the four conserved Cys residues forming the structure of the putative Zn fingers of the carlavirus 3' ORF proteins is replaced with His in the ShVX 15K protein (Fig. 6). Analogous 'finger' structures are found in other plant virus-specific proteins (Sehnke

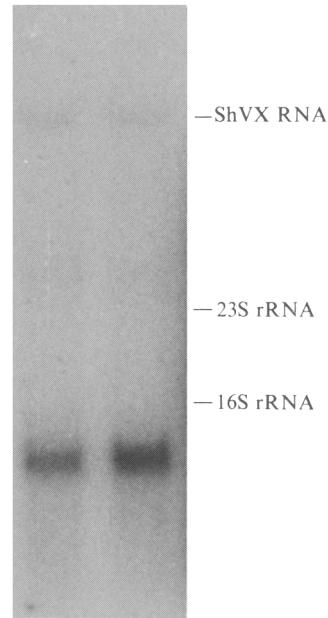


Fig. 7. Hybridization of total RNA, isolated from two individual ShVX-infected shallot plants, with a ³²P-labelled probe complementary to the 3'-terminal region of the ShVX RNA. The positions of marker RNAs are shown.

et al., 1989) and in the case of PVM and CVB have been shown to bind nucleic acids *in vitro* (Gramstat *et al.*, 1990; K. E. Levay & S. K. Zavriev, unpublished observation).

Subgenomic (sg) RNA analysis

By analogy to carla- and potexviruses, the 3'-proximal part of the ShVX genome may be expressed through two sgRNAs (Guilford & Forster, 1986; Dolja *et al.*, 1987; Mackie *et al.*, 1988; Monis & de Zoeten, 1990). To test this hypothesis, we have analysed the total RNA isolated from infected plants by Northern blot hybridization with a ³²P-labelled RNA transcript complementary to the 3'-terminal region of the ShVX RNA (see Methods). Besides genomic RNA, there was only one sgRNA detected, about 1500 nucleotides long (Fig. 7), probably encoding the viral coat protein. The origin of the minor band above the 23S rRNA marker is obscure, and its size (about 4500 nucleotides) makes it an unlikely candidate for the role of a sgRNA for the triple gene block. The absence of a 'conventional' sgRNA about 3500 nucleotides long for the latter is rather surprising, and cannot be adequately explained as yet.

General conclusions

Our analysis of the ShVX genome structure demonstrates that it contains all elements common to carla- and

potexviruses. However, ShVX is distinguished by an unusual gene, ORF4, the product of which has no analogues known to date. Furthermore, a peculiarity of the structural organization of ShVX RNA, like that of LVX (Memelink *et al.*, 1990), is the lack of an ORF for the smaller protein of the triple gene block. It has been shown recently that the triple gene block proteins, including the 7K protein of the potexvirus WCIMV, are required for transport (Beck *et al.*, 1991). Therefore, the lack of analogues of the 7K to 8K proteins in ShVX and LVX can be expected to result in some peculiarities of their transport mechanisms.

Thus, the ShVX genome offers a good example of the evolutionary combination of virus-specific elements now found in different groups of viruses. It can be stated that phylogenetically this virus occupies an intermediate position between carla- and potexviruses.

We would like to thank Drs S. Morozov, T. Konareva and A. Galkin for helpful discussions.

References

- BECK, D. L., GUILFORD, P. J., VOOT, D. M., ANDERSEN, M. T. & FORSTER, R. L. S. (1991). Triple gene block proteins of white clover mosaic potexvirus are required for transport. *Virology* **183**, 695–702.
- BUNDIN, V. S., VISHNYAKOVA, O. A., ZAKHARIEV, V. M., MOROZOV, S. YU., ATABEKOV, J. G. & SKRYABIN, K. G. (1986). Comparative studies of potexvirus genomes: homology between the primary structure of coat protein genes. *Doklady Akademii Nauk SSSR* **290**, 728–733 (in Russian).
- CORPET, F. (1988). Multiple sequence alignment with hierarchical clustering. *Nucleic Acids Research* **16**, 10881–10890.
- DOLJA, V. V., GRAMA, D. P., MOROZOV, S. YU. & ATABEKOV, J. G. (1987). Potato virus X-related single- and double-stranded RNA. Characterization of terminal structures. *FEBS Letters* **214**, 308–312.
- FORSTER, R. L. S., BEVAN, M. W., HARBISON, S. A. & GARDNER, R. C. (1988). The complete nucleotide sequence of the potexvirus white clover mosaic virus. *Nucleic Acids Research* **16**, 291–303.
- FOSTER, G. D., MILLAR, A. W., MEEHAN, B. M. & MILLS, P. R. (1990). Nucleotide sequence of 3'-terminal region of *Helenium* virus S RNA. *Journal of General Virology* **71**, 1877–1880.
- GERMAN, S., CANDRESSE, T., LANNEAU, M., HUET, J. C., PERNOLLET, J. C. & DUNEZ, J. (1990). Nucleotide sequence and genomic organization of apple chlorotic leaf spot virus. *Virology* **179**, 104–112.
- GORBALENYA, A. E., KOONIN, E. V., DONCHENKO, A. P. & BLINOV, V. M. (1988). A novel superfamily of nucleoside triphosphate-binding motif-containing proteins which are probably involved in duplex unwinding in DNA and RNA replication and recombination. *FEBS Letters* **235**, 16–24.
- GRAMSTAT, A., COURTOZANIS, A. & ROHDE, W. (1990). The 12 kDa protein of potato virus M displays properties of a nucleic acid-binding regulatory protein. *FEBS Letters* **276**, 34–38.
- GUILFORD, P. G. & FORSTER, R. L. S. (1986). Detection of polyadenylated subgenomic RNAs in leaves infected with the potexvirus daphne virus X. *Journal of General Virology* **67**, 83–90.
- HABILI, N. & SYMONS, H. (1989). Evolutionary relationship between luteoviruses and other RNA plant viruses based on sequence motifs in their putative RNA polymerases and nucleic acid helicases. *Nucleic Acids Research* **17**, 9543–9555.
- HAYLOR, M. T. M., BRUNT, A. A. & COUTTS, R. H. A. (1990). Conservation of the 3' terminal nucleotide sequence in five carlaviruses. *Nucleic Acids Research* **18**, 6127.
- HOLLINGS, M. & BRUNT, A. A. (1981). Potyviruses. In *Handbook of Plant Virus Infections and Comparative Diagnosis*, pp. 731–807. Edited by E. Kurstak. Amsterdam: Elsevier.
- HUISMAN, M. J., LINTHORST, H. J. M., BOL, J. F. & CORNELISSEN, B. J. C. (1988). The complete nucleotide sequence of potato virus X and its homologies at the amino acid level with various plus-stranded RNA viruses. *Journal of General Virology* **69**, 1789–1798.
- JELKMANN, W., MARTIN, R. R., LESEMANN, D.-E., VETTEN, H. J. & SKELTON, F. (1990). A new potexvirus associated with strawberry mild yellow edge disease. *Journal of General Virology* **71**, 1251–1258.
- KLUG, A. & RHODES, D. (1987). 'Zinc fingers': a novel protein motif for nucleic acid recognition. *Trends in Biochemical Sciences* **12**, 464–469.
- KROCZEK, R. A. & SIEBERT, E. (1990). Optimization of Northern analysis by vacuum-blotting, RNA-transfer visualization, and ultraviolet fixation. *Analytical Biochemistry* **184**, 90–95.
- KYTE, J. & DOOLITTLE, R. F. (1982). A simple method for displaying the hydropathic character of a protein. *Journal of Molecular Biology* **157**, 105–132.
- LAEMMLI, U. K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature, London* **227**, 680–685.
- LEVAY, K. & ZAVRIEV, S. (1991). Nucleotide sequence and gene organization of the 3'-terminal region of chrysanthemum virus B genomic RNA. *Journal of General Virology* **72**, 2333–2337.
- MACKENZIE, D. J., TREMAINE, J. H. & STACE-SMITH, R. (1989). Organization and interterminal homologies of the 3'-terminal portion of potato virus S RNA. *Journal of General Virology* **70**, 1053–1063.
- MACKIE, G. A., JOHNSON, R. & BANCROFT, J. B. (1988). Single- and double-stranded viral RNAs in plants infected with the potexvirus papaya mosaic virus and foxtail mosaic virus. *Intervirology* **29**, 170–177.
- MEMELINK, J., VAN DER VLUGT, C. I. M., LINTHORST, H. J. M., DERKS, A. F. L. M., ASJES, C. J. & BOL, J. F. (1990). Homologies between the genomes of a carlavirus (lily symptomless virus) and potexvirus (lily virus X) from lily plants. *Journal of General Virology* **71**, 917–924.
- MONIS, J. & DE ZOETEN, G. A. (1990). Molecular cloning and physical mapping of potato virus S complementary DNA. *Phytopathology* **80**, 446–450.
- MORCH, M.-D., BOYER, J.-C. & HAENNI, A.-L. (1988). Overlapping open reading frames revealed by complete nucleotide sequencing of turnip yellow mosaic virus genomic RNA. *Nucleic Acids Research* **16**, 6157–6173.
- MOROZOV, S. YU., GORBULEV, V. G., NOVIKOV, V. K., AGRANOVSKI, A. A., KOZLOV, YU. V., ATABEKOV, J. G. & BAYEV, A. A. (1983). The primary structure of the 5' and 3' terminal regions of the genomic RNA of potato virus X. *Doklady Akademii Nauk, SSSR* **259**, 723–725 (in Russian).
- MOROZOV, S. YU., DOLJA, V. V. & ATABEKOV, J. G. (1989). Probable reassortment of genomic elements among elongated RNA-containing plant viruses. *Journal of Molecular Evolution* **29**, 52–62.
- MOROZOV, S. YU., KANYUKA, K. V., LEVAY, K. E. & ZAVRIEV, S. K. (1990a). The putative RNA replicase of potato virus M: obvious sequence similarity with potex- and tymoviruses. *Virology* **179**, 911–914.
- MOROZOV, S. YU., MIROSHNICHENKO, N. A., ZELENINA, D. A., FEDORKIN, O. N., SOLOVJIEV, A. G., LUKASHEVA, L. I. & ATABEKOV, J. G. (1990b). Expression of RNA transcripts of potato virus X full-length and subgenomic cDNAs. *Biochimie* **72**, 677–684.
- MOROZOV, S. YU., MIROSHNICHENKO, N. A., SOLOVJIEV, A. G., ZELENINA, D. A., FEDORKIN, O. N., LUKASHEVA, L. I., GRACHEV, S. A. & CHERNOV, B. K. (1991). *In vitro* membrane binding of the translation products of the carlavirus 7-kDa protein genes. *Virology* **183**, 782–785.
- PALMITER, R. D. (1974). Magnesium precipitation of ribonucleoprotein complexes: expedient techniques for the isolation of undegraded polysomes and messenger ribonucleic acid. *Biochemistry* **13**, 3606–3612.

- POCH, O., SAUVAGET, I., DELARUE, M. & TORDO, N. (1989). Identification of four conserved motifs among the RNA-dependent polymerase encoding elements. *EMBO Journal* **8**, 3867–3874.
- RANDLES, J. M. & ROHDE, W. (1990). *Nicotiana velutina* mosaic virus: evidence for a bipartite genome comprising 3 kb and 8 kb RNAs. *Journal of General Virology* **71**, 1019–1027.
- ROZANOV, M. N., MOROZOV, S. YU. & SKRYABIN, K. G. (1990). Unexpected close relationship between the large nonvirion proteins of filamentous potexviruses and spherical tymoviruses. *Virus Genes* **3**, 370–379.
- RUPASOV, V. V., MOROZOV, S. YU., KANYUKA, K. V. & ZAVRIEV, S. K. (1989). Partial nucleotide sequence of potato virus M RNA shows similarities to potexviruses in gene arrangement and the encoded amino acid sequences. *Journal of General Virology* **70**, 1861–1869.
- SEHNKE, P. C., MASON, A. M., HOOD, S. J., LISTER, R. M. & JOHNSON, J. E. (1989). A 'zinc finger'-type binding domain in tobacco streak virus coat protein. *Virology* **168**, 48–56.
- SIT, T. L., ABOUHAIKAR, M. G. & HOLY, S. (1989). Nucleotide sequence of papaya mosaic virus RNA. *Journal of General Virology* **70**, 2325–2331.
- SIT, T. L., WHITE, K. A., HOLY, S., PADMANABHAN, U., EWEIDA, M., HIEBERT, M., MACKIE, G. A. & ABOUHAIKAR, M. G. (1990). Complete nucleotide sequence of clover yellow mosaic virus RNA. *Journal of General Virology* **71**, 1913–1920.
- SKRYABIN, K. G., MOROZOV, S. YU., KRAEV, A. S., ROZANOV, M. N., CHERNOV, B. K., LUKASHEVA, L. I. & ATABEKOV, J. G. (1988). Conserved and variable elements in RNA genomes of potexviruses. *FEBS Letters* **240**, 33–40.
- TRIFONOV, E. N. (1987). Translation framing code and frame-monitoring mechanism as suggested by the analysis of mRNA and 16S rRNA nucleotide sequences. *Journal of Molecular Biology* **194**, 643–652.
- VISHNICHENKO, V. K., KONAREVA, T. N. & ZAVRIEV, S. K. (1992). A new filamentous virus in shallot. *Plant Pathology* (in press).
- ZAVRIEV, S. K., KANYUKA, K. V. & LEVAY, K. E. (1991). The genome organization of potato virus M RNA. *Journal of General Virology* **72**, 9–14.
- ZUIDEMA, D., LINTHORST, H. J. M., HUISMAN, M. J., ASJES, C. J. & BOL, J. F. (1989). Nucleotide sequence of narcissus mosaic virus RNA. *Journal of General Virology* **70**, 267–276.

(Received 31 March 1992; Accepted 4 June 1992)